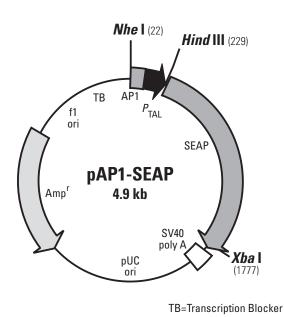
pAP1-SEAP Vector Information

GenBank Accession No.: Submission in progress.



Restriction Map of pAP1-SEAP. All restriction sites are unique.

Description:

pAP1-SEAP is designed for monitoring induction of the activator protein 1 (AP1; 1) and the stress-activated protein kinase/Jun N-terminal kinase (SAPK/JNK) signal transduction pathway (2). pAP1-SEAP contains the secreted alkaline phosphatase (SEAP) reporter gene (3–5). This vector also contains four tandem copies of the AP1 enhancer fused to a TATA-like promoter (P_{TAL}) region from the Herpes simplex virus thymidine kinase (HSV-TK) promoter. After transcription factors bind AP1, transcription is induced and the reporter gene is activated.

The SEAP coding sequence is followed by the SV40 late polyadenylation signal to ensure proper, efficient processing of the SEAP transcript in eukaryotic cells. Located upstream of AP1 is a synthetic transcription blocker (TB), which is composed of adjacent polyadenylation and transcription pause sites for reducing background transcription (6). The vector backbone also contains an f1 origin for single-stranded DNA production, a pUC origin of replication, and an ampicillin resistance gene for propagation and selection in *E. coli*.





United States/Canada 800.662.2566 Asia Pacific +1.650.919.7300 Europe +33.(0)1.3904.6880 Japan +81.(0)77.543.6116

Clontech Laboratories, Inc. ATakara Bio Company 1290 Terra Bella Ave. Mountain View, CA 94043 Technical Support (US) E-mail: tech@clontech.com www.clontech.com

Use:

pAP1-SEAP is designed to measure the binding of transcription factors to AP1, providing a direct measurement of activation for this pathway. For example, the addition of serum or growth factors to the cell-culture medium induces the binding of transcription factors to AP1, which initiates transcription of SEAP. Alternatively, you can cotransfect this vector with an expression vector containing your gene of interest to monitor pathway activation. The secreted SEAP enzyme can be assayed directly from the culture medium using one of Clontech's Great EscAPe Chemiluminescence Detection Kits (Cat. Nos. 631701, 631704). In addition, the SEAP assay permits time-course studies not possible with assays dependent on cell lysates. The pAP1-SEAP Vectors can be transfected into mammalian cells by any standard method. For selecting stable clones, cotransfect with a vector containing an antibiotic resistance gene, such as neomycin, hygromycin, or puromycin.

(PR93783; published 05 May 1999)

Location of features:

- Activator Protein 1 (AP1; 1) element: 27–72
- TATA-like promoter (*P*_{τΔ1}): 79–227
- Secreted alkaline phosphatase (SEAP) gene:
 - SEAP coding sequences: start codon (ATG): 255-257; stop codon: 1812-1814 signal peptide: 255-305 mature protein: 306-1811 C-terminal extension to SEAP: 1773-1811
- SV40 late mRNA polyadenylation signal: 1925–1930 mRNA 3' end: 1944
- pUC plasmid replication origin: 2323–2966
- Ampicillin resistance gene:

Promoter: -35 region: 4044-4039; -10 region: 4021-4016

Transcription start point: 4009

Ribosome binding site: 3986-3982

β-lactamase coding sequences:

start codon (ATG): 3974-3972; stop codon: 3116-3114

β-lactamase signal peptide: 3974–3906

β-lactamase mature protein: 3905–3117

- f1 single-strand DNA origin (packages the noncoding strand of SEAP): 4106–4561
- Transcription blocker (TB): 4692–4845

Synthetic polyadenylation site (7): 4692-4740

Transcription pause site from human $\alpha 2$ globin gene (8): 4754–4845

Propagation in *E. coli*:

- Suitable host strains: DH5 α and other general purpose strains. Single-stranded DNA production requires a host containing an F' episome such as JM109.
- Selectable marker: plasmid confers resistance to ampicillin (50 µg/ml) to E. coli hosts.
- E. coli replication origin: pUC
- Copy number: ~500
- Plasmid incompatibility group: pMB1/Col E1

References:

- Latchman, D. S. (1995) In Eukaryotic Transcription Factors, 2nd Ed. (Academic Press, San Diego), pp. 72. 1.
- Cahill, M. A., et al. (1996) Curr. Biol. 6:16-19. 2
- Yang, T. T., et al. (July 1994) CLONTECHniques IX(3):1-5. 3. Berger, J., et al. (1988) Gene 66:1-10. 4.
- 5.
- Cullen, B. R. & Malim, M. H. (1992) Methods Enzymol. 216:362-368. Eggermont, J. & Proudfoot, N. (1993) EMBO J. 12:2539-2548. 6.
- Levitt, N., et al. (1989) Genes Dev. 3:1019-1025. 7.
- 8. Enriquez-Harris, P., et al. (1991) EMBO J. 10:1833-1842.

Notice to Purchaser

The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech. This vector has not been completely sequenced.

Clontech products are to be used for research purposes only. They may not be used for any other purpose, including, but not limited to, use in drugs, in vitro diagnostic purposes, therapeutics, or in humans. Clontech products may not be transferred to third parties, resold, modified for resale, or used to manufacture commercial products or to provide a service to third parties without written approval of Clontech Laboratories, Inc

Clontech, the Clontech logo and all other trademarks are the property of Clontech Laboratories, Inc. Clontech is a Takara Bio Company. ©2006